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## EXPRESSION OF DYSTROPHIN-ASSOCIATED GLYCOPROTEINS DURING HUMAN FETAL MUSCLE DEVELOPMENT: A PRELIMINARY IMMUNOCYTOCHEMICAL STUDY

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**Abstract**-An immunocytochemical study was performed on quadriceps muscle from eight fetuses ranging from 12 weeks of gestation to term, using antibodies against the dystrophin-associated proteins, in order to evaluate the developmental expression of these proteins. For comparison, antibodies against dystrophin and utrophin were also used. The expression of the 59 kDa dystrophin-associated protein was simultaneous with that of dystrophin, which is also a subsarcolemmal protein. The extracellular glycoprotein of 156 kDa ( $\alpha$ -dystroglycan) and the transmembrane glycoprotein of 43 kDa ( $\beta$ -dystroglycan) appeared to be expressed later. The transmembrane glycoproteins of 50 kDa (adhalin) and 35 kDa were fully expressed at an even later stage of fetal muscle development. This study suggests that the subsarcolemmal proteins may have an essential role in the assembly of the transmembrane and extracellular components of the dystrophin-glycoprotein complex during fetal muscle development. The knowledge obtained from observing the developmental expression of these proteins may contribute to the understanding of the molecular mechanism of their different involvement in muscle disorders.

Human fetal muscle, development, dystrophin, utrophin, dystrophin-glycoprotein complex, dystrophin-associated proteins, dystroglycan, adhalin.

### INTRODUCTION

The discovery of a large oligomeric complex of glycoproteins associated with dystrophin gave a new insight to the understanding of the molecular mechanism of Duchenne and Becker muscular dystrophies [1-6]. This complex spans the sarcolemma providing a link between the subsarcolemmal cytoskeleton and laminin, a major component of the extracellular matrix [a-9]. The complex is composed of four glycoproteins (156, 50, 43 and 35 kDa) and two proteins (59 and 25 kDa). The 156 and 43 kDa glycoproteins (named  $\alpha$ -dystroglycan and  $\beta$ -dystroglycan, respectively) are encoded by a single gene mapped to chromosome 3q21[9]. The  $\alpha$ -dystroglycan is an extracellular glycoprotein which binds to laminin and is linked to

$\beta$ -dystroglycan, a transmembrane protein [10]. The 50 and 35 kDa glycoproteins and the 25 kDa protein are also transmembrane while the 59 kDa protein is cytoplasmic and is linked to dystrophin". In Duchenne muscular dystrophy (DMD), the absence of dystrophin leads to a great reduction in dystrophin-associated proteins, thus disrupting the linkage between the subsarcolemmal cytoskeleton and the extracellular matrix. This is presumed to render muscle fibers susceptible to necrosis. In Becker muscular dystrophy (BMD) it was recently shown that the partial deficiency of dystrophin correlates both in intensity and distribution with the reduction of all of the dystrophin-associated proteins [1, 12].

The 50 kDa dystrophin-associated glycoprotein (adhalin), is a novel transmembrane glycoprotein [13] expressed almost exclusively in skeletal and cardiac muscles [13, 14]. Adhalin is deficient in severe childhood autosomal recessive muscular dystrophy with DMD-like phenotype

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(SCARMD) [15], a disease first reported in Tunisia [16] and prevalent in North African countries, which has been linked to chromosome 13q12 [17, 18]. Adhalin deficiency was subsequently found in European patients [19] and it was recently demonstrated that SCARMD is genetically heterogeneous, as some Brazilian and French families with adhalin deficiency do not map to chromosome 13 [20, 21].

The modifications of the dystrophin-glycoprotein complex and its role in the pathogenesis of muscular dystrophies have recently been discussed in several publications [10, 22-24]. Here is presented a preliminary study of the distribution and localization of the proteins of the dystrophin-glycoprotein complex during the development of human fetal muscle.

#### MATERIALS AND METHODS

Muscle samples from quadriceps muscle were obtained under legal conditions from eight fetuses ranging from 12 weeks of gestation to term. The age of the fetuses was determined using different well established criteria [25-27]. The muscle samples were collected shortly after delivery and they were frozen in isopentane cooled in liquid nitrogen and stored at  $-80^{\circ}\text{C}$ . Conventional histological and histochemical techniques were performed on 10  $\mu\text{m}$  transverse cryostat sections of frozen muscle specimens, as in other studies [28]. This allowed us to ascertain the state of preservation of the muscle tissue and its degree of differentiation. This study was carried out only in fetuses not presenting either autolytic or pathological changes.

For immunocytochemistry, 7  $\mu\text{m}$  transverse cryostat sections of muscle specimens from fetuses of different ages and from an infant of 9 months were placed on superfrost glass microscope slides and processed as previously described [5, 6, 11-12, 15, 29]. Indirect immunofluorescence was performed using antibodies against dystrophin (directed against the C-terminal, rod and N-terminal domains), utrophin, the 59 kDa dystrophin-associated protein (59DAP), and the 156 kDa (156DAG or a-dystroglycan), 50 kDa (SODAG or adhalin), 43 kDa (43DAG or pdystroglycan) and 35 kDa (35DAG) dystrophin-associated glycoproteins, previously characterized [5, 6, 11-12, 15, 29]. Other monoclonal anti-dystrophin antibodies (NLC DYS 2 and NLC DYS 3, Novocastra) were also used. The sections were examined under a Zeiss AxioPlan fluorescence microscope

and the photographs were taken under identical conditions with the same exposure time.

#### RESULTS AND DISCUSSION

The antibodies against dystrophin and utrophin showed an expression of these proteins in the fetus of 12 weeks of gestational age (Fig. 1), as has been reported in other studies [3&35]. The expression of these two proteins diverged during subsequent stages of fetal muscle development: dystrophin progressively increased until birth while utrophin rapidly decreased. At 12 weeks of gestation, the antibody against utrophin showed strong labelling of the periphery of the myotubes, but at 32 weeks the labelling of the sarcolemma of the muscle cells was already faint. At birth it was almost absent but remained prominent at the neuromuscular junctions, as was also seen in the 9-month-old child.

The first protein of the dystrophin-associated complex to be abundantly expressed was the 59DAP. In fetuses of 12 weeks of development the antibody against this protein labelled the periphery of myotubes (Fig. 1). This labelling was discontinuous in most muscle cells but in older fetuses it became progressively more regular and at 32 weeks was clearly distinct around the periphery of nearly all muscle fibers. The labelling with the antibody against the 59DAP during development was concomitant with that obtained with the anti-dystrophin antibodies (Fig. 1).

The 156DAG and 43DAG ( $\alpha$  and  $\beta$ -dystroglycan, respectively) were little expressed in the younger fetuses (Fig. 2). Their expression became more apparent in fetuses of 16 weeks of development and in older fetuses it progressively increased in intensity. At 32 weeks most fibers were already labelled by the antibodies against these glycoproteins.

The expression of adhalin and 35DAG was delayed in relation to that of  $\alpha$ - and  $\beta$ -dystroglycan (Fig. 3). Until the age of 22 weeks very few muscle cells were labelled by the antibodies against these two glycoproteins, particularly with the antibody against adhalin. At 32 weeks the labelling with the antibodies against adhalin and 35DAG was still irregular and less intense than that seen at term and in the 9-month-old infant.

The neuromuscular junctions were labelled with the antibodies against all proteins of the dystrophin-glycoprotein complex. The labelling of neuromuscular junctions is illustrated in

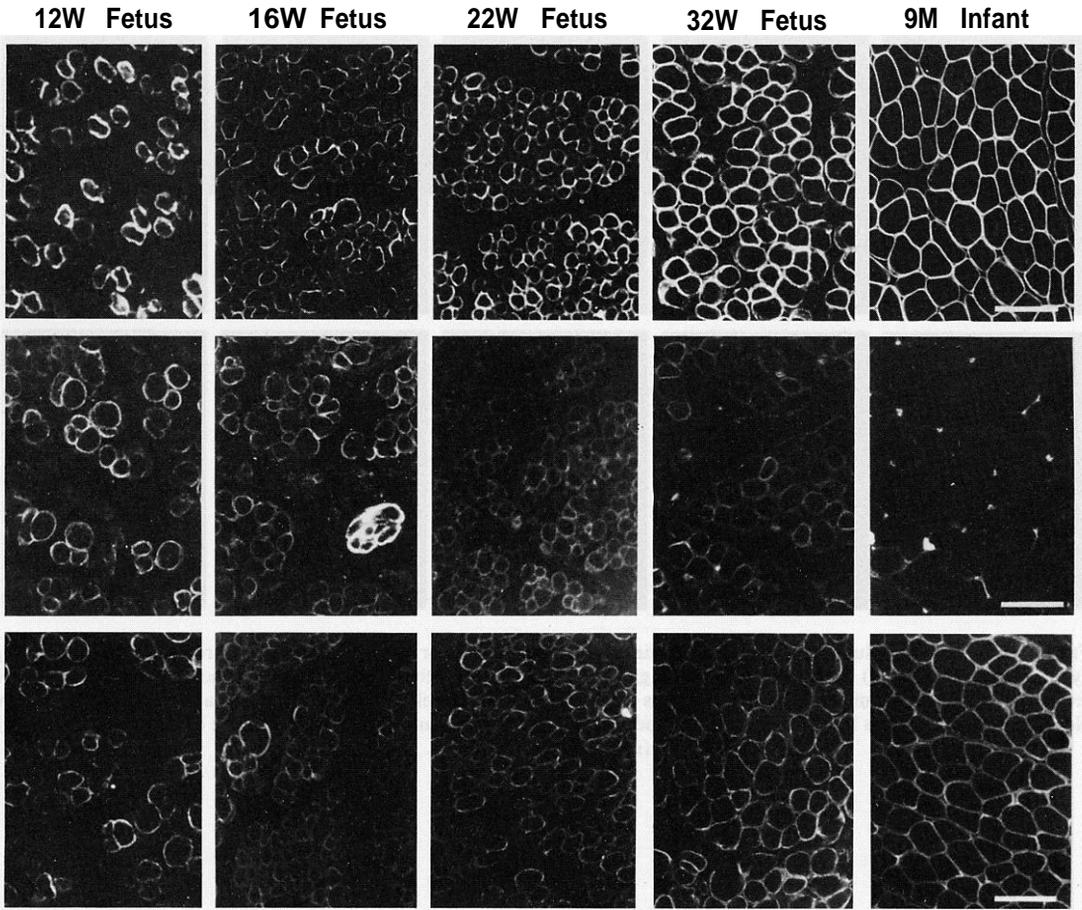


FIG. 1. Indirect immunofluorescence with antibodies against the C-terminal domain of dystrophin (upper row), utrophin (middle row) and the 59 kDa dystrophin-associated protein (lower row) showing the expression of these proteins in cryostat transverse sections of quadriceps muscle from human fetuses of 12, 16, 22 and 32 weeks of development and a child aged nine months. Bar 20  $\mu\text{m}$ .

the 9-month-old child with antibodies against – dystroglycan (Fig. 2) and adhalin (Fig. 3). No labelling with any of the antibodies used was seen in cells at the myoblast stage, lying between the myotubes in the younger fetuses.

The results of this study are schematically illustrated in Fig. 4. The strong expression of utrophin at an early age of fetal development has been reported previously [33-35]. Dystrophin and 59DAP are also expressed early but at a lower rate. These three proteins are subsarcolemmal and their expression appears to precede the expression of the transmembrane and extracellular glycoproteins of the dystrophin-glycoprotein complex. This suggests that they may play an essential role in the assembly of the glycoproteins of the dystrophin-glycoprotein complex. The concomitant expression of both the – and – dystroglycan correlates with the fact that these two glycoproteins are closely related and have a common messenger RNA [6, 9]. The concomi-

tant expression of adhalin and 35DAG may also suggest a link between the two glycoproteins. This may explain why in SCARMD, a condition characterized by a deficiency in adhalin, there is a slight reduction in 35DAG [15, 19]. It is interesting to note that the drastic decrease in utrophin occurs in fetuses of about 22-24 weeks of gestation and this is concomitant with the installation of monoaxonal innervation [36] and muscle fiber type cyto-enzymological differentiation [37, 38]. Utrophin persists at the neuromuscular junction in Duchenne muscular dystrophy and *mdx* mice [39-41], and may compensate for dystrophin deficiency [42, 43]. It appears to be nerve dependent, as it is overexpressed after experimental denervation in *mdx* mice [44, 45].

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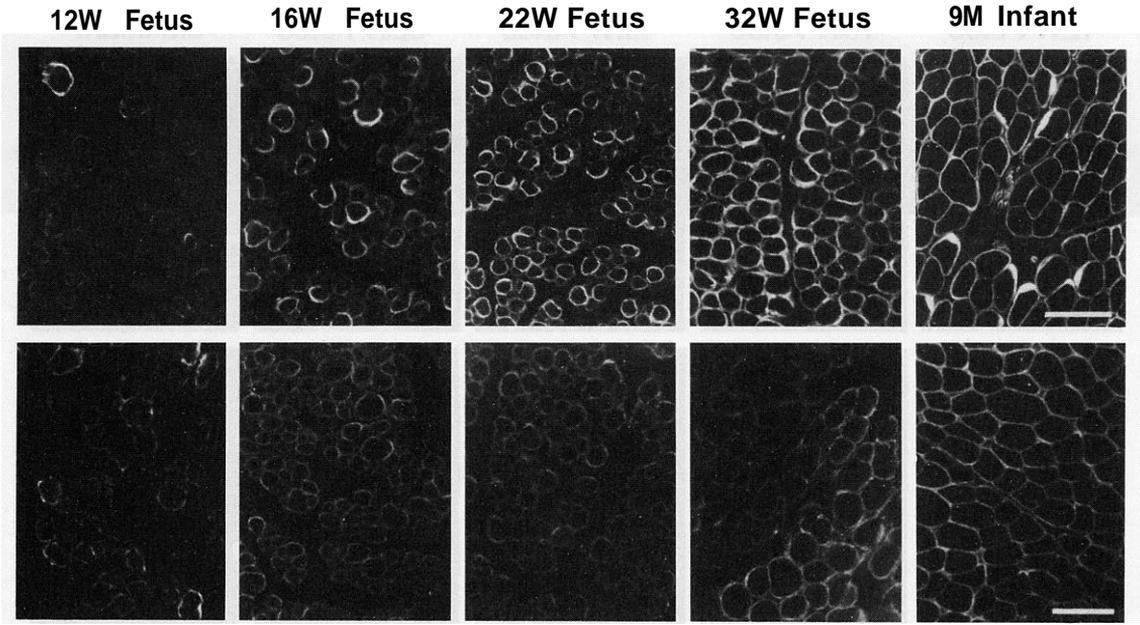


FIG. 2. Indirect immunofluorescence with antibodies against the 156 kDa or dystroglycan (upper row) and the 43 kDa or  $\beta$ -dystroglycan (lower row) dystrophin-associated glycoproteins, showing their progressive expression in cryostat transverse sections of quadriceps muscle from human fetuses of 12, 16, 22 and 32 weeks of development and a child aged nine months. Neuromuscular junctions labelled with the antibody against  $\alpha$ -dystroglycan are seen in the muscle of the 9-month-old child (upper row). Bar 20  $\mu$ m.

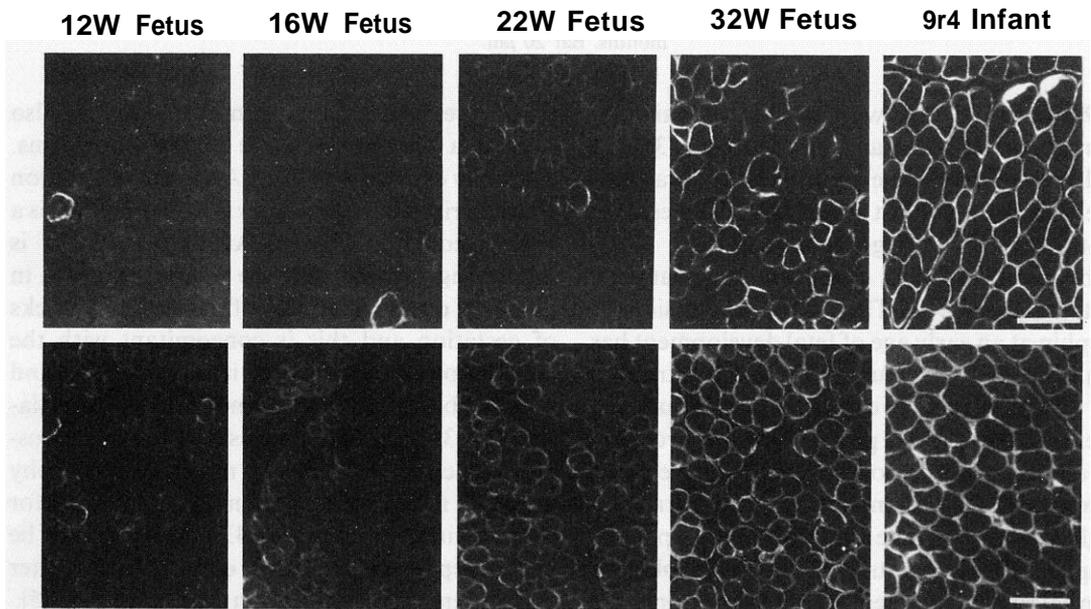


FIG. 3. Indirect immunofluorescence with antibodies against the 50 kDa (adhalin) (upper row) and 35 kDa dystrophin-associated glycoproteins (lower row), showing their expression in cryostat transverse sections of quadriceps muscle from human fetuses of 12, 16, 22 and 32 weeks of development and a child aged nine months. Neuromuscular junctions labelled with the antibody against adhalin are seen in the muscle of the 9-month-old child (upper row). Bar 20  $\mu$ m.

## Developmental Expression of Dystrophin, Utrophin and Dystrophin-Associated Proteins in Human Skeletal Muscle

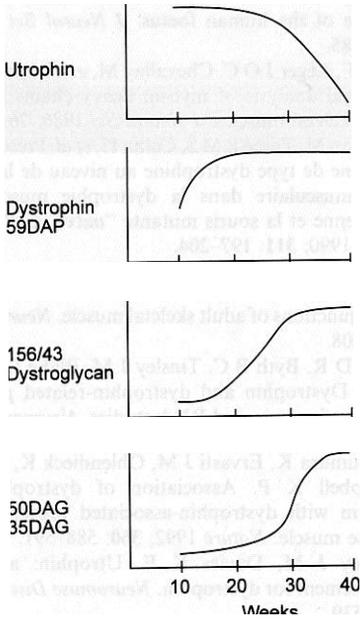


FIG. 4. Schematic representation of the sequential expression of utrophin, dystrophin and dystrophin-associated proteins during human fetal muscle development.

## REFERENCES

- Campbell K P, Kahl D S. Association of dystrophin and integral membrane glycoprotein. *Nature* 1989; 338: 259-262.
- Ervasti J M, Dblendieck K, Kahl S D, et al. Deficiency of a glycoprotein component of the dystrophin-glycoprotein complex in dystrophic muscle. *Nature* 1990; 345: 315-319.
- Yoshida M, Dzawa E. Glycoprotein complex anchoring dystrophin to sarcolemma. *J Biochem (Tokyo)* 1990; 108: 748-752.
- Ohlndieck K, Ervasti J M, Snook J B, Campbell K P. Dystrophin-glycoprotein complex is highly enriched in isolated skeletal muscle sarcolemma. *J Cell Bio*, 199; 112: 135-148.
- Ervasti J M, Campbell K P. Membrane organization of the dystrophin-glycoprotein complex. *Cell*, 1991; 66: 1121-1131.
- Ibraghimov-Beskrovnaya O, Ervasti I M, Lweille C J, et al. Primary structure of dystrophin-associated glycoproteins linking dystrophin to extracellular matrix. *Nature* 1992; 355: m-702.
- Ervasti J M, Campbell K P. A role for the dystrophin-glycoprotein complex as a transmembrane linker between laminin and actin. *J Cell Biol* 1993; 122: 809-823.
- Gee S H, Blather R W, Douville P J, et al. Laminin-binding protein 120 from brain is closely related to the dystrophin-associated glycoprotein, dystroglycan, and binds with high affinity to the major heparin binding domain of laminin. *J Biol Chem* 1993; 268: 14,972-14,980.
- Ibraghimov-Beskrovnaya O, Milatovich A, Dzelic T, et al. Human dystroglycan: skeletal muscle cDNA, genomic structure, origin of tissue specific forms and chromosomal localization. *Hum Mol Genet* 1993; 2: 1651-1657.
- Ervasti J M, Campbell K P. Dystrophin-associated glycoproteins: their possible roles in the pathogenesis of Duchenne muscular dystrophy. In: Partridge T, ed. *Molecular and Cell Biology of Muscular Dystrophy*. London: Chapman & Hall, 1993: 139-166.
- Matsumura K, Nonaka I, Tom & F M S, et al. Mild deficiency of dystrophin-associated proteins in Becker muscular dystrophy patients having in-frame deletions in the rod domain of dystrophin. *Am J Hum Genet* 1993; 53: 409-416.
- Matsumura K, Burghes A H M, Mom M, et al. Immunocytochemical analysis of dystrophin-associated proteins in Becker/Duchenne muscular with huge in-frame deletions of N-terminal and rod domains of dystrophin. *J Clin Invest* 1994; 93: 99-105.
- Raberds S L, Anderson R D, Ibraghimov-Beskrovnaya O, Campbell K P. Primary structure and muscle-specific expression of the 50 kDa dystrophin-associated glycoprotein (adhain). *J Biochem* 1993; 268: 23,739-23,742.
- Yamamoto H, Mizuno Y, Hayashi K, Nonaka I, Yoshida M, Dzawa E. Expression of dystrophin-associated protein 35DAG (A4) and SODAG (A2) is confined to striated muscles. *J Biochem (Tokyo)*, 1994; 115: 162-167.
- Matsumura K, Tom & F M S, Collin H; et al. Deficiency of the 50K dystrophin-associated glycoprotein in severe childhood autosomal recessive muscular dystrophy. *Nature* 1992; 359: 320-322.
- Ben Hamida M, Fardeau M, Attia N. Severe childhood muscular dystrophy affecting both sexes and frequent in Tunisia. *Muscle Nerve* 1983; 6: 469-480.
- Ben Dthmane K, Ben Hamida M, Pericak-Vance M A, et al. Linkage of Tunisian autosomal recessive Duchenne-like muscular dystrophy to the pericentromerit region of chromosome 13q. *Nature Genet* 1992; 2: 35-37.
- Azibi K, Bachner L, Beckman J S, et al. Severe childhood autosomal recessive muscular dystrophy with the deficiency of the 50 kDa dystrophin-associated glycoprotein maps to chromosome 13q12. *Hum Mol Genet* 1993; 2: 1423-1428.
- Fardeau M, Matsumura K, Tom & F M S, et al. Deficiency of the 50 kDa dystrophin-associated glycoprotein (adhain) in severe childhood autosomal recessive muscular dystrophies in children native from European countries. *C R Acad Sci Paris* 1993; 316: 799-804.
- Passes-Buena M R, Oliveira J R, Bakker E, et al. Genetic heterogeneity for Duchenne-like muscular dystrophy (DLMD) based on linkage and 50 DAG analysis. *Hum Mol Genet* 1993; 2: 1945-1947.
- Romero N B, Tom & F M S, Leturcq F, et al. Genetic heterogeneity of severe childhood autosomal recessive muscular dystrophy with adhain (50 kDa dystrophin associated glycoprotein) deficiency. *CR Acad Sci Paris* 1994; 317: 7&K.
- Matsumura K, Campbell K P. Deficiency of dystrophin-associated proteins: a common mechanism leading to muscle cell necrosis in severe childhood muscular dystrophies. *Neuromuscul Disord* 1993; 3: 109-118.
- Matsumura K, Ohlndieck K, Ionasexu V V, et al. The role of the dystrophin-glycoprotein complex in the molecular pathogenesis of muscular dystrophies. *Neuromuscul Disord* 1993; 3: 533-535.
- Matsumura K, Campbell K P. Dystrophin-glycoprotein complex: its role in the molecular pathogenesis of muscular dystrophies. *Muscle Nerve*, 1994; 17: 2-15.
- Scammon R E, Calkins L A. *The Development and Growth of the External Dimensions of the Human Body in the Fetal Period*. Minneapolis: The University of Minneapolis Press, 1929.
- Trotti D. Age of fetuses determined from their increases. *Acta Obstet Gynec Scand* 1948; 27: 327-337.

27. Hamilton W J, Mossman H W. Human ~mb,yo,0~,, 4th edn. Cambridge: H&r & Sons, ,972.

28. Fardeau M. Caractéristiques cytochimiques et ultra-structurales des différents types de fibres musculaires squelettiques entrafusales (chez l'homme et quelques mammifères). Ann Anar Palhol 1973; 18: 7-34.

29. Ohlendicck K, Matsumura K, Ionasescu V V, et ol. Duchenne muscular dystrophy: deficiency of dystrophin-associated proteins in the sarcolemma. Neuro,ogy 1993; 43: 795-800.

30. Wessels A, Ginjaar I B, Maorman A F M, van Ommen G J B. Different localization of dystrophin in developing and adult human skeletal muscle. Muscle Nerve 1991; 14: 1-11.

31. Prella A, Chianese L, Moggio M, et ol. Appearance and localization of dystrophin in normal human fetal m"SCK I", J De" Neurosc 1991; 9: 607-612.

32. Clerk A, Strong P N, Sewry C A. Characterisation of dyrtrophin during development of human skeletal muscle. Deve/opmen~ 1992; 114: 3951102.

33. Clerk A, Morris G E, Dubowitz V, Davies K E, Sewry C A. Dystrophin-related protein, utrophin, in normal and dystrophic human fetal skeletal muscle. Hisrochem J 1993; 25: 556561.

34. Khurana T S, Watkins S C, Chafey P, et a., Immunolocalization and developmental expression of dystrophin related protein. JCCnBiol1991,115(3)part 3: 178a.

35. Tom& F M S, Chevally M, Leturcq F, et al. Immunocytochemical and biochemical study of dystrophin and dystrophin-related protein (DRP) during development of skeletal muscle in normal, and DMD fetuses. Neurology 1992, 42 (suppl. 3): 227-228.

36. Tom6 F M S, Rieger F, Chevally M, e, a., Early stages

of the development of human muscle fibers and their innervation. Muscle Nerve 1986; 9(5S): 173.

37. Coling-Saltin A S. Enzyme histochemistry on skeletal muscle of the human foetus. J Neural Sci 1978; 39: 169-185.

38. Pons F, L&r J O C, Chevally M, e, a., hnmunocyto-chemical analysis of myosin heavy chains in human fetal skeletal muscles. J Neuro, Sci ,986: 76, 151-163.

39. FardeauM,TombFM S, CollinH, era., Pr&enced'une prothe de type dystrophine au niveau de la jonction neuromusculaire dans la dystrophie musculaire de Duchenne et la souris mutante "mdx". C R Aead Sci Pôris 1990; 311: 197-204.

40. Ohlendieck K, Ervasti J M, Matsumura K, ef ol. Dystrophin-related protein is localized to neuromuscular junctions ofadult skeletal muscle. Neuron 1991; 7: 499-508.

41. Love D R, Byth B C, Tinsley J M, Blake D I, Davies K E. Dystrophin and dystrophin-related proteins: a review of protein and RNA studies. Neuromuse LXX& 1993; 3: 5-21.

a. Matsumura K, Ervasti J M, Ohlendieck K, Kahl S D, Campbell K P. Association of dystrophin-related protein with dystrophin-assoCiated proteins in mdx mouse muscle. Narure 1992; 360: 588-591.

43. Tinsley J M, Davies K E. Utrophin: a potential replacement fordystrophin. NeuromurcD~sord 1993; 3: 537-539.

44. Takemitsu M, Ishiura S, Koga R, ef al. Dystrophin-related protein in the fetal and denervated skeletal muscles of normal and mdx mice. Biochem Biophys Res Commun 1991; 180: 1179-1186.

45. Alameddine H, Tomb F M S (to be published).

REFERENCES

1. Campbell K P, Kahl D S. Association of dystrophin and utrophin with the sarcolemma. Nature 1987; 328: 289-291.

2. Ervasti J M, Ohlendieck K, Kahl S D, et al. Dystrophin-associated protein complex of the dystrophin-related protein in dystrophic muscle. Nature 1990; 345: 312-319.

3. Yoshida M, Ozawa E. Glycoprotein complex anchoring dystrophin to sarcolemma. J Biochem (Tokyo) 1990; 108: 744-752.

4. Ohlendieck K, Ervasti J M, Snow B, Campbell K P. Dystrophin-glycoprotein complex is highly enriched in isolated skeletal muscle sarcolemma. J Cell Biol 1991; 111: 145-148.

5. Ervasti J M, Campbell K P. Membrane organization of the dystrophin-glycoprotein complex. Cell 1991; 66: 1131-1137.

6. Dragatsis-Barkovaya O, Ervasti J M, Laville C J, et al. Primary structure of dystrophin-associated glycoprotein linking dystrophin to sarcolemma. Nature 1992; 358: 598-602.

7. Ervasti J M, Campbell K P. A role for the dystrophin-glycoprotein complex as a transmembrane linker between laminin and actin. J Cell Biol 1992; 122: 509-522.

8. Got S H, Blacketer R W, Dowdell P J, et al. Laminin-binding protein 120 from brain is closely related to the dystrophin-associated glycoprotein dystroglycan and binds with high affinity to the major heparin-binding domain of laminin. J Biol Chem 1992; 267: 14922-14930.

9. Dragatsis-Barkovaya O, Mitsuhashi A, Otsuki T, et al. Human dystroglycan skeletal muscle cDNA: genome structure, origin of tissue specific isoforms and chromosomal localization. Hum Mol Genet 1992; 1: 1451-1457.

10. Ervasti J M, Campbell K P. Dystrophin-associated

11. Atfeh K, Rachez L, Beckmann J S, et al. Severe childhood autosomal recessive muscular dystrophy with the deletion of the 50 kDa dystrophin-associated glycoprotein maps to chromosome 13q12. Hum Mol Genet 1992; 1: 143-147.

12. Fardeau M, Matsumura K, Tomé F M S, et al. Evidence of the 50kDa dystrophin-associated glycoprotein (beta-DG) in severe childhood autosomal protein (beta-dystrophin) in children native from recessive muscular dystrophy in children native from European countries. C R Acad Sci Paris 1992; 314: 799-804.

13. Fardeau M, Rachez L, Rachez J R, Dierker E, et al. Genetic heterogeneity for Duchenne-like muscular dystrophy (DMD) based on linkage and 50 kDa dystrophin-associated glycoprotein (beta-DG) analysis. Hum Mol Genet 1992; 1: 1947-1947.

14. Rieger F, Ervasti J M, Tomé F M S, et al. Genetic heterogeneity of severe childhood autosomal recessive muscular dystrophy with absence 50 kDa dystrophin-associated glycoprotein. C R Acad Sci Paris 1994; 317: 30-30.

15. Matsumura K, Campbell K P. Dystrophin and dystrophin-associated protein: a normal heparin-binding protein with cell adhesion in severe childhood muscular dystrophy. Neurosci Biomed 1992; 3: 109-114.

16. Matsumura K, Ohlendieck K, Matsumura Y, et al. The role of the dystrophin-glycoprotein complex in the molecular pathogenesis of muscular dystrophies. Neurosci Biomed 1992; 3: 221-221.

17. Matsumura K, Campbell K P. Dystrophin-glycoprotein complex: its role in the molecular pathogenesis of muscular dystrophies. Trends Neurosci 1992; 15: 1-12.

18. Scamman E E, Carlson J A. The biochemistry and genetics of the X-linked Duchenne of the human form in the fetal period. Minneapolis: The University of Minnesota Press 1978.

19. Teff D. A new method for determining the amino acid sequence of a protein. J Biol Chem 1971; 246: 1457-1462.