

Matters Arising

The Naming of Voltage-Gated Calcium Channels

Voltage-gated calcium channels are multisubunit complexes formed of a central channel-forming α_1 subunit and several regulatory and/or auxiliary subunits which include a β subunit and the disulfide-linked $\alpha_2\delta$ subunit. Depending on the tissue of origin, a fifth subunit, such as the skeletal muscle γ or the neuronal p95, may also form part of the channel complex. Additional subunits may still be discovered. Molecular cloning has greatly expanded the understanding of calcium channel diversity, but confusion remains in the naming of the plethora of genes that encode calcium channel α_1 and β subunits. This is further complicated by the fact that the transcripts of most subunits are subject to alternative splicing, which in some but not all cases, may be

tissue specific. Table 1 compares the names of cloned α_1 genes and their mature mRNAs, their major sites of known expression, and their functional correlates as they are perceived today. Table 2 lists β subunit genes splice variants, and some sites of expression.

To simplify matters, the undersigned propose to use a unified nomenclature based on rules that allow the description of a mature assembled heteromeric channel in terms of an $\alpha_{1X}\beta_n\gamma_n\delta_n$ complex, where X is a capital letter (S, A, B, C, D, E, etc.) that identifies the genes from which the α_1 subunit originates, and n is a number (1, 2, 3, etc.) that identifies the genes from which other calcium channel subunits originate. α_{1S} denotes the α_1 subunit of the skeletal muscle calcium channel, the first of these channels to be cloned in the laboratory of the late Professor Shosaku Numa. α_{1A} through α_{1E} denote the α_1 subunits cloned subsequently, labeled with an

Table 1. Calcium Channel α_1 Subunits

| Gene Product | | Functional Correlates | | |
|-------------------|---|---|--------------------------------|---|
| Consensus Name(s) | Original Name(s) if Different | Sites of Expression | Current | Drug Sensitivity of Native Currents |
| α_{1S} | Skeletal muscle CaCh1 <i>0.1skm</i> | Skeletal muscle, BC3H1 cells | HVA L type | Sensitive to DHPs, diltiazem and verapamil Insensitive to sub- μ M ω -CTx-GVIA and funnel web spider venoms (ω -Aga-IVA, FTX) |
| α_{1A} | BI CaCh4 rbA | Brain, cerebellum, Purkinje and granule cells, kidney, PC12 cells, C cells | HVA Q type? HVA P type? | ω -CTx-MVIIC (>100 nM); ω -CTx-G DHP insensitive Sensitive to ω -Aga-IVA (<10nM) and low sFTX DHP insensitive |
| α_{1B} | BIII CaCh5 rbB | Brain, peripheral neurons, PC12 cells, C cells | HVA N type | Sensitive to ω -CTx-GVIA (100-500 nM) and ω -CTx-MVIIC (>100 nM) DHP insensitive |
| α_{1C} | Cardiac Smooth muscle/ lung CaCh2 rbC | Heart, HIT cells, GH3 cells, brain, aorta, lung, kidney, fibroblasts PC12 cells, C cells | HVA L type | DHP sensitive Insensitive to low concentrations of ω -CTx-GVIA, ω -Aga-IVA, or sFTX |
| α_{1C-a} | CaCh2a | Heart | | |
| α_{1C-b} | CaCh2-l CaCh2b CaCh2-II | Smooth muscle, lung | | |
| α_{1C-c} | rbC CaCh2-III | Brain | | |
| α_{1D} | CaCh3 Neuroendocrine rbD | Brain, pancreas, HIT cells, GH3 cells, PC12 cells, C cells | HVA L type | DHP sensitive Reversibly sensitive to ω -CTx-GVIA, ω -Aga-IVA, or FTX |
| α_{1E} | CaCh6 BII rbE | Brain, heart, C cells | HVA R type? | Sensitive to low Ni Insensitive to DHPs or ω -CTx-MVIIC, or to low concentrations of ω -CTx-GVIA, ω -Aga-IVA or sFTX |

This table is intended as a guide and refers only to mammalian calcium channels. Not all previously used names are listed. Vertebrate *doe-1* and *doe-4* α_1 subunits, cloned from the marine ray *Discopyge ommata*, are orthologs of mammalian α_{1E} and α_{1B} , respectively. HVA and LVA, high and low voltage activated; DHP, dihydropyridine; ω -CTx-G and ω -CTx-M, ω -conotoxins from marine snails *Conus geographus* and *Conus magus*, respectively; Aga, agatoxin (funnel web spider *Agelenopsis aperta* toxin); sFTX, synthetic funnel web spider toxin. Q-type calcium channel: current in cerebellar granule cells sensitive to ω -CTx-MVIIC but insensitive to DHPs, low ω -CTx-GVIA, and low ω -Aga-IVA; R-type calcium channel: residual in cerebellar granule cells after blocking with DHP, ω -Aga-IVA, ω -CTx-GVIA, and ω -CTx_MVIIC.

Table 2. Calcium Channel β Subunits

| Gene Product | Splice Variant | Other Name(s) | Proven Expression ^a | Component of |
|--------------|----------------|------------------------|--------------------------------|------------------------|
| β_1 | β_{1a} | β_{1M} | Skeletal muscle | DHP receptor |
| | β_{1b} | β_{1B2}, β_2 | Brain, heart | ? |
| | β_{1c} | β_{1B1} | Brain, heart | ? |
| β_2 | β_{2a} | β_3 | Brain, heart | ? |
| | β_{2b} | | Brain, heart | ? |
| | β_{2c} | | Brain, heart | ? |
| β_3 | ? | | Brain, heart, aorta | ω -CTx receptor |
| β_4 | ? | | Brain | ? |

^aDoes not exclude sites of expression.

empirical terminology developed for the calcium channels from brain, the only tissue in which all of these genes are expressed. The genes encoding regulatory/auxiliary subunits (β , $\alpha_2\delta$, γ , etc.) are numbered sequentially in approximate order of their discovery. Note that a single $\alpha_2\delta$ gene and mRNA yields two mature subunits, α_2 and δ , which are disulfide linked. Thus, the $\alpha_2\delta_1$ to $\alpha_2\delta_n$ genes are expected to encode a series of disulfide-linked subunit pairs (α_2 and δ) in the mature calcium channel protein.

Splice variants are uniformly denoted by y , a lower-case letter (i.e., α_{1A-a} , α_{1A-b} , β_{1a} , β_{1b} , $\alpha_{2\delta a}$, $\alpha_{2\delta b}$, etc.). If no second gene is known, such as for the $\alpha_2\delta$ subunit, the capital letter or numerical subscript is omitted. If no molecular diversity is known, such as for the γ subunit of the skeletal muscle calcium channel, subscripts are omitted. In this nomenclature, the skeletal muscle L-type calcium channel/dihydropyridine receptor has the subunit composition $\alpha_{1S}\beta_{1a}\gamma\alpha_{2\delta a}$.

Signed (alphabetical listing):

Lutz Birnbaumer

Department of Anesthesiology
UCLA School of Medicine
Los Angeles, California 90024

Kevin P. Campbell

Howard Hughes Medical Institute
and Department of Physiology and Biophysics
University of Iowa School of Medicine
Iowa City, Iowa 52242

William A. Catterall

Department of Pharmacology
University of Washington School of Medicine
Seattle, Washington 98195

Michael M. Harpold

The Salk Institute

Biotechnology/Industrial Associates, Inc. (SIBIA)
La Jolla, California 92037

Franz Hofmann

Institute of Pharmacology and Toxicology
Technical University of Munich
Munich 40 80802
Federal Republic of Germany

William A. Horne

Department of Molecular and Cellular Physiology
Stanford University School of Medicine
Stanford, California 94305

Yasuo Mori

Institute of Pharmacology and Cell Biophysics
University of Cincinnati School of Medicine
Cincinnati, Ohio 45267

Arnold Schwartz

Institute of Pharmacology and Cell Biophysics
University of Cincinnati School of Medicine
Cincinnati, Ohio 45267

Terry P. Snutch

Departments of Zoology and Neurosciences
Biotechnology Laboratory
University of British Columbia
Vancouver
Canada V6T 1Z3

Tsutomu Tanabe

Howard Hughes Medical Institute
and Department of Cellular and Molecular Physiology
Yale University School of Medicine
New Haven, Connecticut 06536

Richard W. Tsien

Department of Molecular and Cellular Physiology
Stanford University School of Medicine
Stanford, California 94305